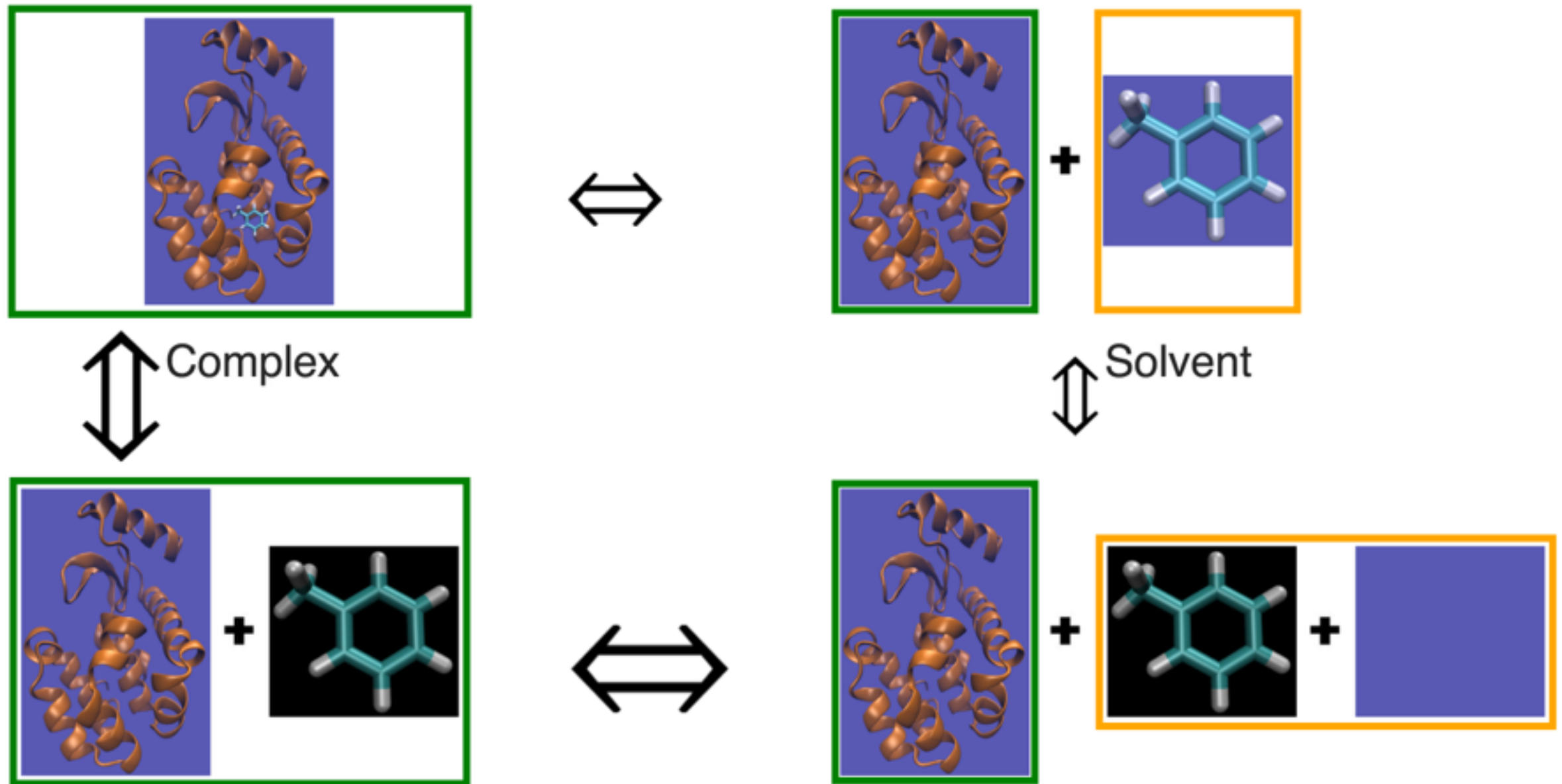


Run free energy calculations with YANK

MSM workshop @ Novartis, Boston - 5/27/2016

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What does YANK do?



0) Install YANK

1. Install miniconda (or anaconda)

<http://conda.pydata.org/miniconda.html>

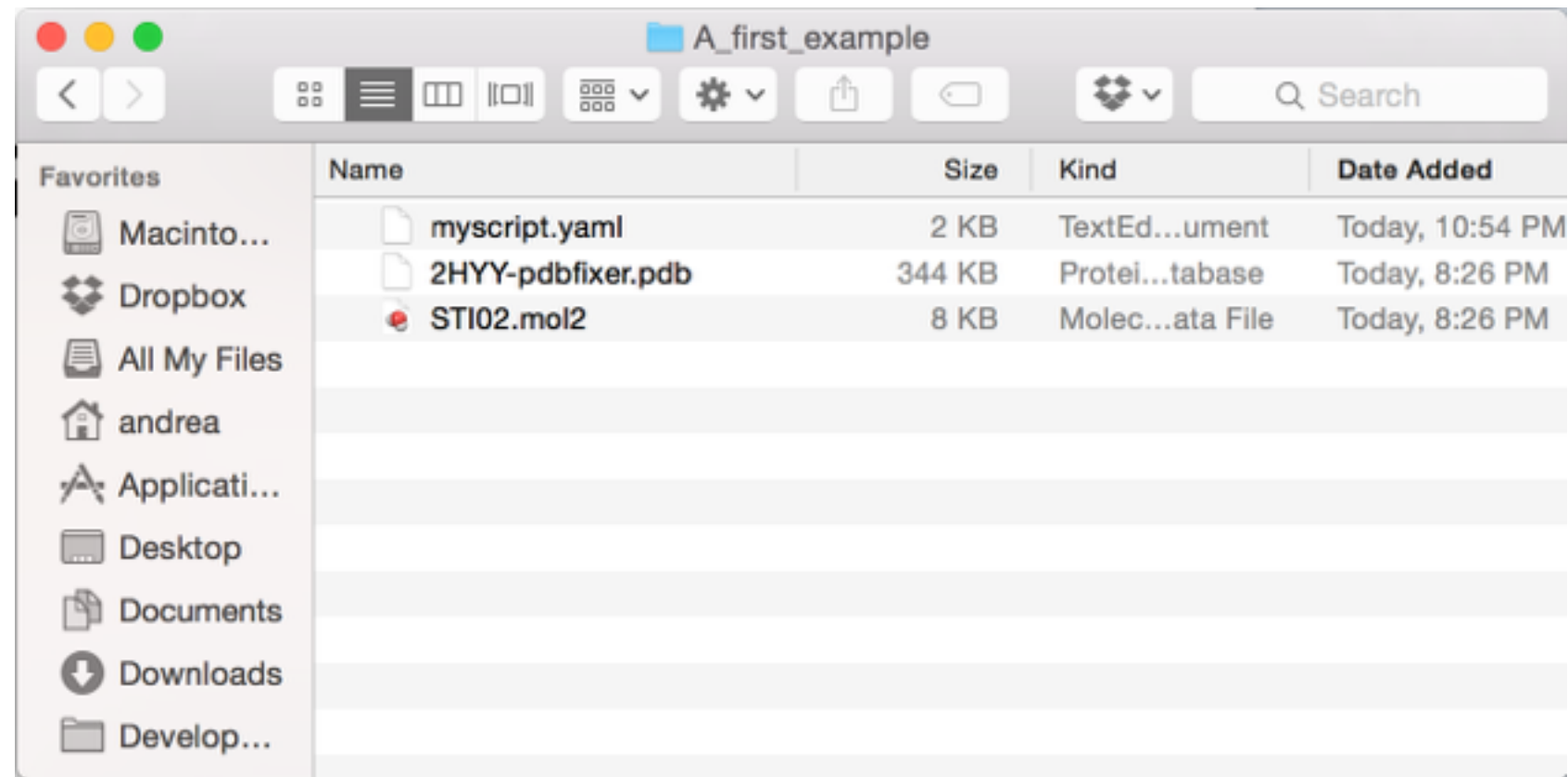
2. Install yank

```
conda config --add channels http://conda.binstar.org/omnia  
conda install yank
```

0.a) Why miniconda?

- It's so easy
- Automatically resolve dependencies
- It doesn't mess up your system
 - Installs everything in your home folder, no admin rights required (i.e. works on clusters)
 - You can create parallel environments to avoid conflicts
- Can deploy essentially anything, not just Python

1) Get all the input files



- YANK configuration file (will see details later)
- Receptor structure
- Ligand structure

2) Run

```
yank script --yaml myscript.yaml
```

Automatically set up system
Hamiltonian Replica Exchange

3) Analyze results

```
yank analyze --store experiment/
```

MBAR

First example: YAML script

```
---  
molecules:  
  Abl:  
    filepath: 2HHY-pdbfixer.pdb  
    parameters: leaprc.ff14SB  
  imatinib:  
    filepath: STI02.mol2  
    parameters: openeye:am1bcc-gaff  
  
solvents:  
  RF: # reaction field  
    nonbondedMethod: CutoffPeriodic  
    nonbondedCutoff: 9*angstroms  
    clearance: 9*angstroms  
    positive_ion: Na+  
    negative_ion: Cl-  
  
experiment:  
  components:  
    receptor: Abl  
    ligand: imatinib  
    solvent: RF  
  protocol: absolute-binding  
  
protocols:  
  absolute-binding:  
    ... # more on this later  
  
options:  
  ... # more on this later
```


First example: YAML script

```
---
molecules:
  Abl:
    filepath: 2HHY-pdbfixer.pdb
    parameters: leaprc.ff14SB
  imatinib:
    filepath: STI02.mol2
    parameters: openeye:am1bcc-gaff

solvents:
  RF: # reaction field
    nonbondedMethod: CutoffPeriodic
    nonbondedCutoff: 9*angstroms
    clearance: 9*angstroms
    positive_ion: Na+
    negative_ion: Cl-

experiment:
  components:
    receptor: Abl
    ligand: imatinib
    solvent: RF
  protocol: absolute-binding

protocols:
  absolute-binding:
    ... # more on this later

options:
  ... # more on this later
```

First example: YAML script

```
---  
molecules:  
  Abl:  
    filepath: 2HHY-pdbfixer.pdb  
    parameters: leaprc.ff14SB  
  imatinib:  
    filepath: STI02.mol2  
    parameters: openeye:am1bcc-gaff
```



```
solvents:  
  RF: # reaction field  
    nonbondedMethod: CutoffPeriodic  
    nonbondedCutoff: 9*angstroms  
    clearance: 9*angstroms  
    positive_ion: Na+  
    negative_ion: Cl-
```

<http://docs.eyesopen.com/toolkits/cookbook/python/modeling/am1-bcc.html>

```
experiment:  
  components:  
    receptor: Abl  
    ligand: imatinib  
    solvent: RF  
  protocol: absolute-binding
```

```
protocols:  
  absolute-binding:  
    ... # more on this later
```

```
options:  
  ... # more on this later
```

First example: YAML script

```
---
molecules:
  Abl:
    filepath: 2HHY-pdbfixer.pdb
    parameters: leaprc.ff14SB
  imatinib:
    filepath: STI02.mol2
    parameters: antechamber

solvents:
  RF: # reaction field
    nonbondedMethod: CutoffPeriodic
    nonbondedCutoff: 9*angstroms
    clearance: 9*angstroms
    positive_ion: Na+
    negative_ion: Cl-

experiment:
  components:
    receptor: Abl
    ligand: imatinib
    solvent: RF
  protocol: absolute-binding

protocols:
  absolute-binding:
    ... # more on this later

options:
  ... # more on this later
```

```
# Parameterize ligand from Tripos mol2.
echo "Parameterizing ligand with GAFF and AM1-BCC charges..."
antechamber -fi mol2 -i pxylene.mol2 -fo mol2 -o pxylene.gaff.mol2 -c bcc
parmchk -i pxylene.gaff.mol2 -o pxylene.gaff.frcmod -f mol2
```

First example: YAML script

```
---  
molecules:  
  Abl:  
    filepath: 2HHY-pdbfixer.pdb  
    parameters: leaprc.ff14SB  
  imatinib:  
    filepath: STI02.mol2  
    parameters: openeye:am1bcc-gaff
```

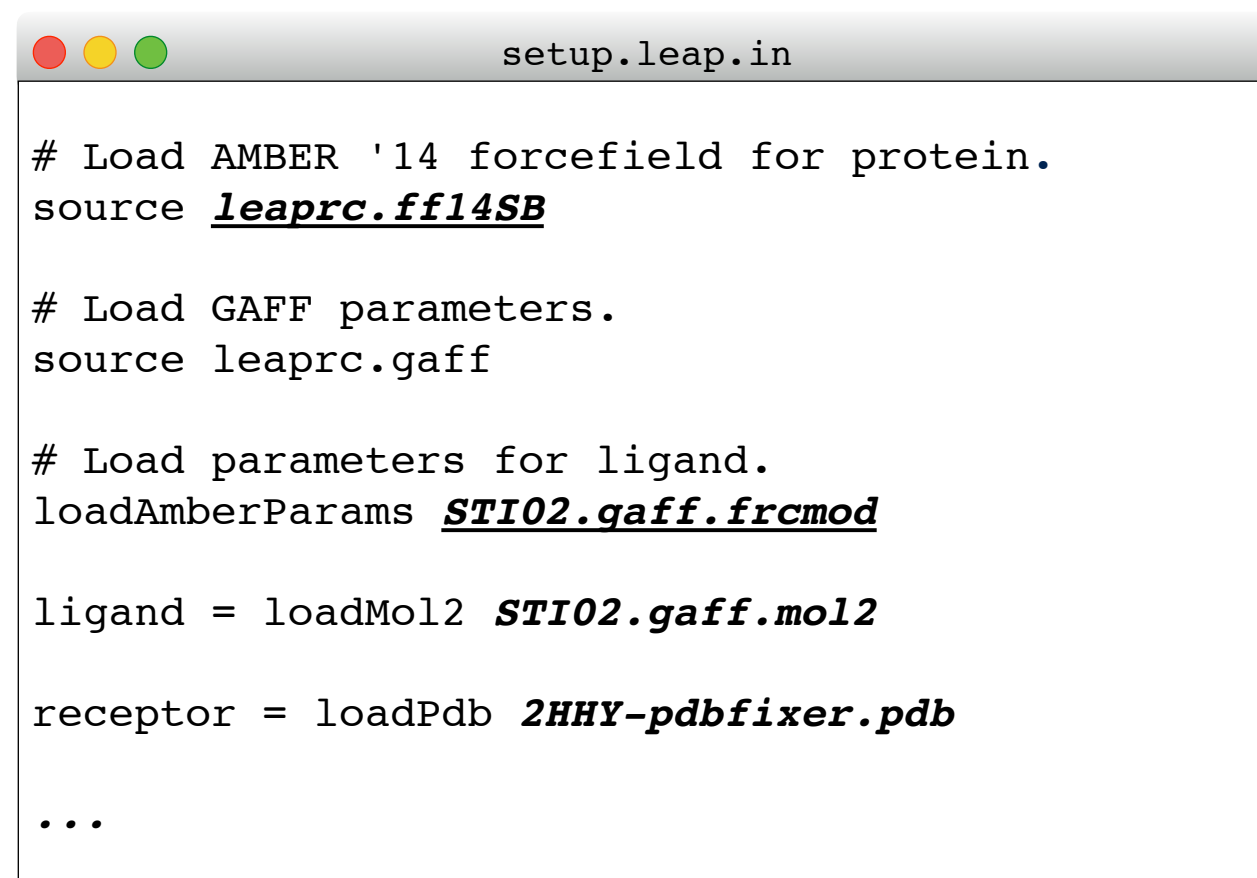
```
solvents:  
  RF: # reaction field  
    nonbondedMethod: CutoffPeriodic  
    nonbondedCutoff: 9*angstroms  
    clearance: 9*angstroms  
    positive_ion: Na+  
    negative_ion: Cl-
```

```
experiment:  
  components:  
    receptor: Abl  
    ligand: imatinib  
    solvent: RF  
  protocol: absolute-binding
```

```
protocols:  
  absolute-binding:  
    ... # more on this later
```

```
options:  
  ... # more on this later
```

```
# Create AMBER prmtop/inpcrd files.  
echo "Creating AMBER prmtop/inpcrd files..."  
tleap -f setup.leap.in > setup.leap.out
```



```
setup.leap.in  
  
# Load AMBER '14 forcefield for protein.  
source leaprc.ff14SB  
  
# Load GAFF parameters.  
source leaprc.gaff  
  
# Load parameters for ligand.  
loadAmberParams STI02.gaff.frcmod  
  
ligand = loadMol2 STI02.gaff.mol2  
  
receptor = loadPdb 2HHY-pdbfixer.pdb  
  
...
```

First example: YAML script

```
---
molecules:
  Abl:
    filepath: 2HHY-pdbfixer.pdb
    parameters: leaprc.ff14SB
  imatinib:
    filepath: STI02.mol2
    parameters: openeye:amlbcc-gaff

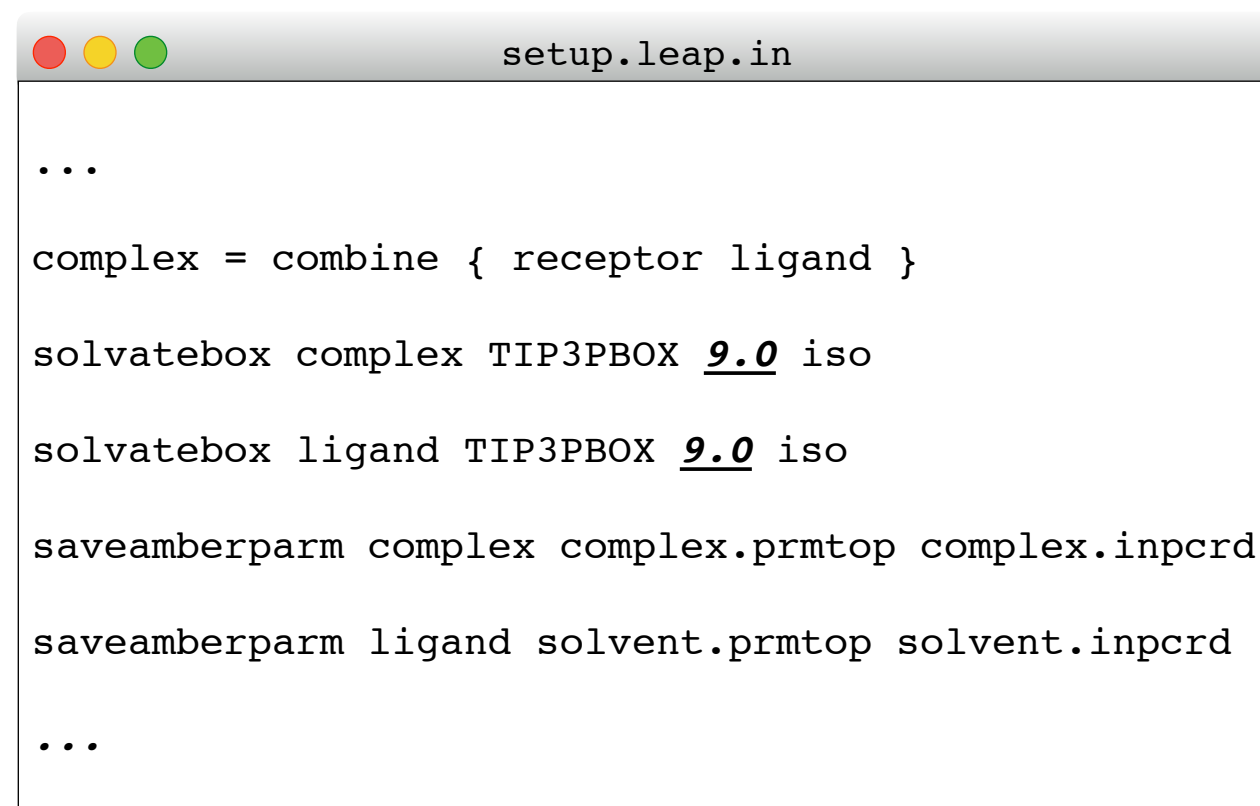
solvents:
  RF: # reaction field
    nonbondedMethod: CutoffPeriodic
    nonbondedCutoff: 9*angstroms
    clearance: 9*angstroms
    positive_ion: Na+
    negative_ion: Cl-

experiment:
  components:
    receptor: Abl
    ligand: imatinib
    solvent: RF
  protocol: absolute-binding

protocols:
  absolute-binding:
    ... # more on this later

options:
  ... # more on this later
```

```
# Create AMBER prmtop/inpcrd files.
echo "Creating AMBER prmtop/inpcrd files..."
tleap -f setup.leap.in > setup.leap.out
```



```
setup.leap.in

...
complex = combine { receptor ligand }
solvatebox complex TIP3PBOX 9.0 iso
solvatebox ligand TIP3PBOX 9.0 iso
saveamberparm complex complex.prmtop complex.inpcrd
saveamberparm ligand solvent.prmtop solvent.inpcrd
...
```

First example: YAML script

```
---
molecules:
  ...

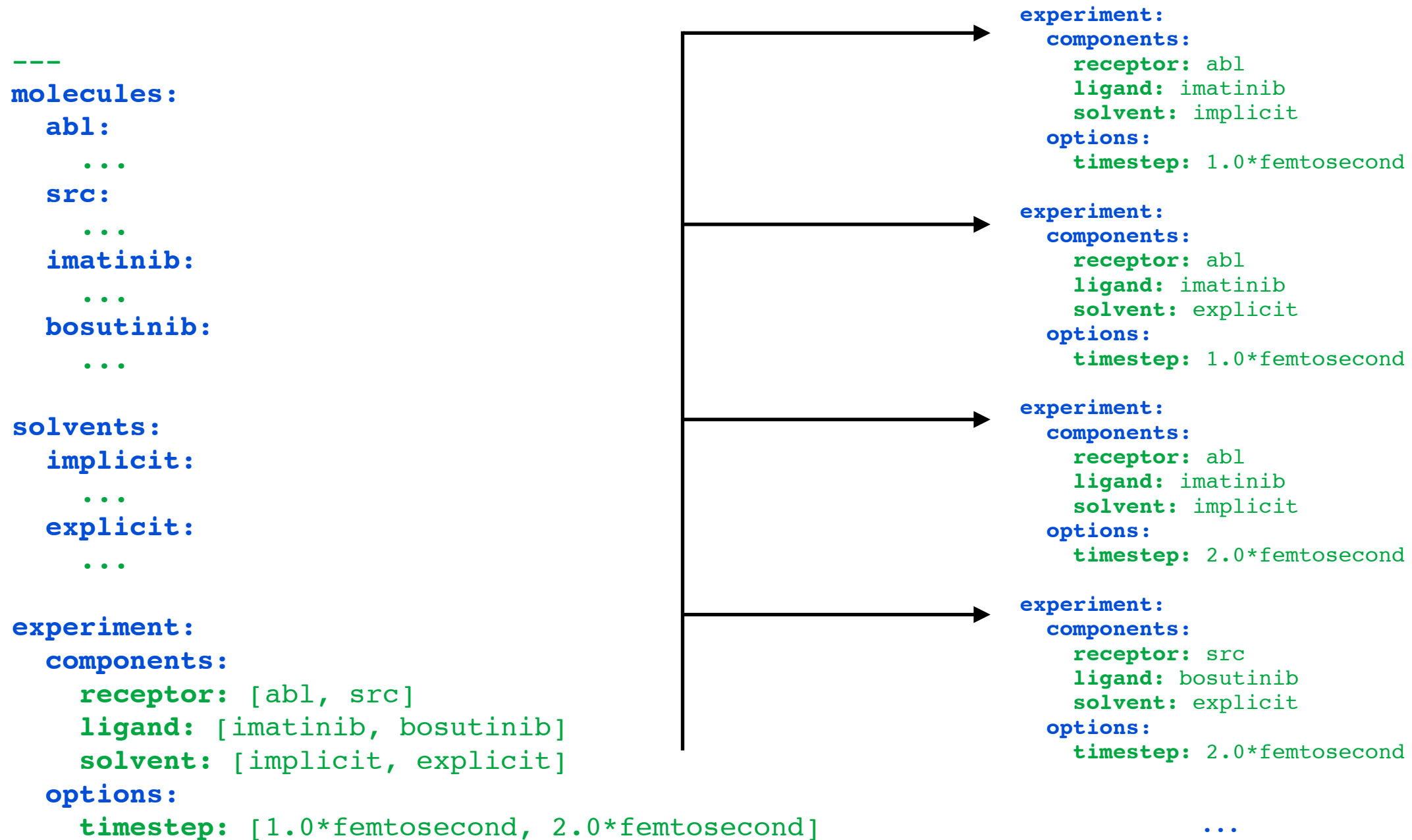
solvents:
  ...

experiment:
  components:
    ...
  protocol: absolute-binding

options:
  minimize: yes
  verbose: yes
  mpi: yes
  number_of_iterations: 1000
  output_dir: ./
  temperature: 300*kelvin
  pressure: 1*atmosphere
  softcore_beta: 0.0

protocols:
  absolute-binding:
    phases:
      complex:
        alchemical_path:
          lambda_electrostatics: [1.0, 0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2, 0.1, 0.0, ...]
          lambda_sterics: [1.0, 1.0, 1.0, 1.0, 1.0, 1.0, 1.0, 1.0, 1.0, 1.0, 0.9, 0.8, ...]
        solvent:
          alchemical_path:
            lambda_electrostatics: [1.0, 0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2, 0.1, 0.0, ...]
            lambda_sterics: [1.0, 1.0, 1.0, 1.0, 1.0, 1.0, 1.0, 1.0, 1.0, 1.0, 0.9, 0.8, ...]
```

Combinations of experiments



More options

```
options:
  verbose: true
  mpi: yes
  resume_setup: true
  resume_simulation: yes
  output_dir: path/to/outputdir
  temperature: 300*kelvins
  pressure: 1*atmosphere
  constraints: AllBonds
  hydrogen_mass: 2*amus # hydrogen mass repartitioning
  restraint_type: harmonic
  randomize_ligand: yes
  randomize_ligand_sigma_multiplier: 2.0 # ligand randomization maximum displacement
  randomize_ligand_close_cutoff: 1.5*angstrom # ligand randomization threshold for overlapping atoms
  collision_rate: 5.0 / picosecond # collision rate for Langevin dynamics
  constraint_tolerance: 1.0e-6
  timestep: 2.0*femtosecond
  nsteps_per_iteration: 2500
  number_of_iteration: 1000
  equilibration_timestep: 1.0*femtosecond
  number_of_equilibration_iterations: 100
  minimize: False
  minimize_tolerance: 1.0 * kilojoules_per_mole / nanometers
  minimize_max_iterations: 100
  replica_mixing_scheme: swap-all

...
```

<https://github.com/choderalab/yank/blob/master/examples/yank-yaml-cookbook/all-options.yaml>

More molecules

molecules:

Abl_combinatorial:

filepath: [2HHY-pdbfixer.pdb, 3CS9-pdbfixer.pdb] # We are working on a protein pipeline

parameters: [leaprc.ff14SB, oldff/leaprc.ff99SBildn]

benzene_sdf:

filepath: benzene.sdf

parameters: antechamber

benzene_smiles:

smiles: c1ccccc1

parameters: antechamber

benzene_name:

name: benzene

parameters: antechamber

benzene_protonation:

name: benzene

parameters: antechamber

epik: 1

More solvents

solvents:

vacuum:

nonbondedMethod: NoCutoff

GBSA-OBC2:

nonbondedMethod: NoCutoff

implicitSolvent: OBC2

PME:

nonbondedMethod: PME

nonbondedCutoff: 1*nanometer

clearance: 10*angstroms

ReactionField:

nonbondedMethod: CutoffNonPeriodic

nonbondedCutoff: 1*nanometer

clearance: 10*angstroms

OpenMM 7.0 Benchmarks

Performance measured in ns/day. [Details.](#)

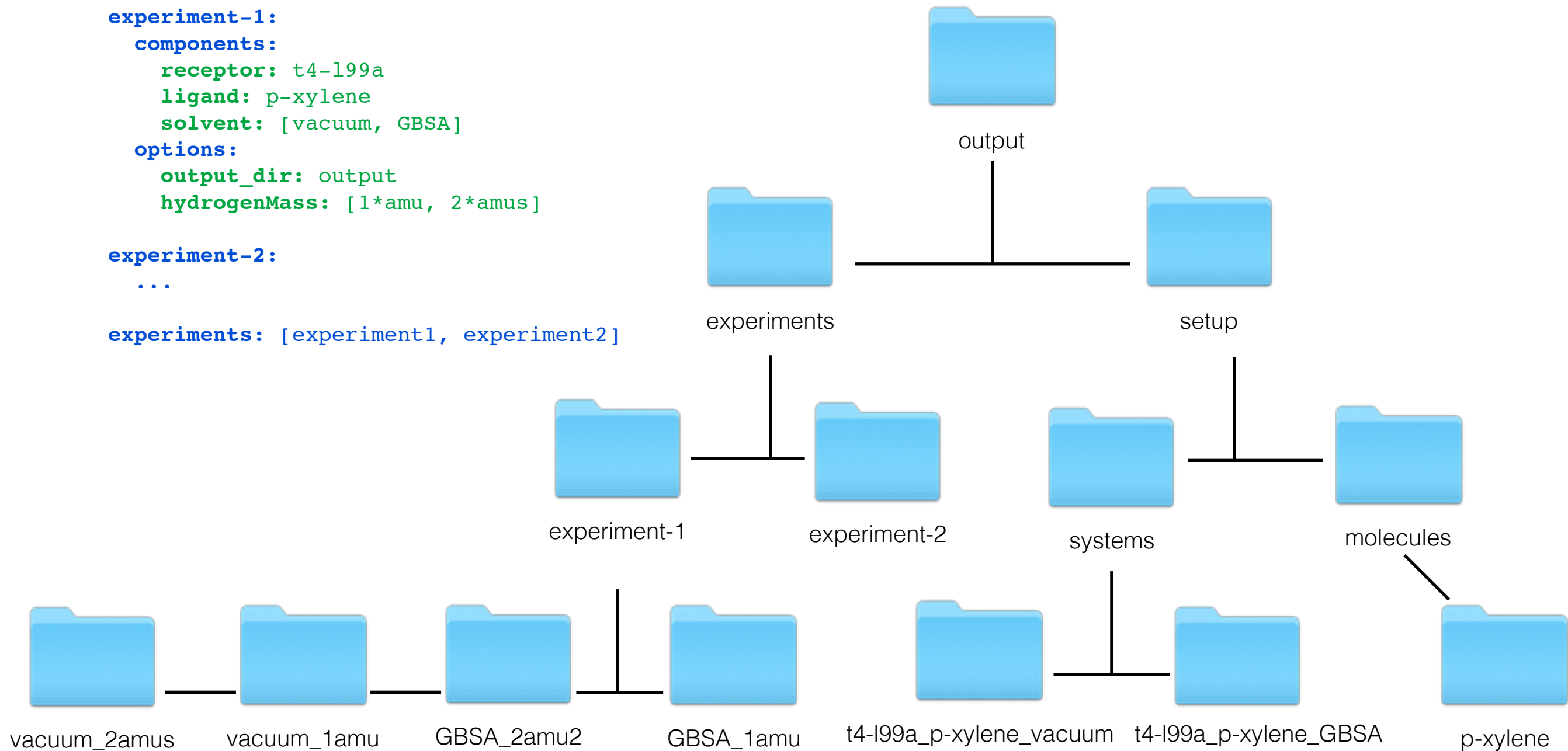
Simulation Type	CUDA (GTX Titan X)	OpenCL (GTX Titan X)	OpenCL (Radeon R9 Fury)	CPU (i7-2700K)
Implicit, 2 fs	482	361	195	8.5
Implicit, 5 fs HMR	756	592	258	20.4
Explicit-RF, 2 fs	309	266	154	16.5
Explicit-RF, 5 fs HMR	542	470	216	38.0
Explicit-PME, 2 fs	204	154	81	12.8
Explicit-PME, 5 fs HMR	415	330	144	30.2

Sequences of experiments

```
experiment-abl:  
  components:  
    receptor: abl  
    ligand: [imatinib, regorafenib, cabozantinib]  
    solvent: [implicit, explicit]  
  options:  
    number_of_iteration: 1000  
  
experiment-src:  
  components:  
    receptor: src  
    ligand: [imatinib, regorafenib, cabozantinib]  
    solvent: [implicit, explicit]  
  options:  
    number_of_iteration: 2000  
  
experiments: [experiment-abl, experiment-src]
```

Output folder structure

```
experiment-1:  
  components:  
    receptor: t4-l99a  
    ligand: p-xylene  
    solvent: [vacuum, GBSA]  
  options:  
    output_dir: output  
    hydrogenMass: [1*amu, 2*amus]  
experiment-2:  
  ...  
experiments: [experiment1, experiment2]
```



Resume

options:

resume_setup: yes

resume_simulation: no

- Initial dry run to check beforehand if the experiment will be interrupted but it still can happen
- YANK does not overwrite anything, you need to remove the files if you want them to be generated again

More docs

- Main docs: <http://getyank.org/latest/>
- Examples: <https://github.com/choderalab/yank/tree/master/examples>

```
yank script skeleton
```

- YAML cookbook: <https://github.com/choderalab/yank/tree/master/examples/yank-yaml-cookbook>
- This presentation: andrea.rizzi@choderalab.org

Near future

- CHARMM force field (Chaya Stern)
- Protein setup automatic pipeline
- Easy movie
- Solvation free energy
- Clustering binding modes (Rosa Luirink, Daan Geerke group in University of Amsterdam)
- Relative free energy calculations (*semi-near* future)